## **Piledriver and 23 Mile Slough Survey Report 2017**

Tanana Valley Watershed Association October 25, 2017



### Introduction

This report discloses the findings of the 2017 study undertook by the Tanana Valley Watershed Association (TVWA) on Piledriver Slough and 23 mile Slough. For this study, Piledriver Slough was dissected into upper and lower subdivisions for monitoring. Monitoring on Upper Piledriver Slough was done with the assistance of citizen student-scientist from Salcha School. Survey site results are discussed below.

### Purpose

This ten-year study is within its sixth year pursuant to fulfillment of the Mitigation Measure 56 of the Service Transportation Board. The measure states, "prior to construction of Salcha Alternative Segment 1, ARRC shall develop appropriate mitigation in consultation with ADF&G to prevent blockage of Piledriver and 23 Mile Sloughs by beaver dams (as a result of flushing flows caused by ARRC-proposed channel plugs). Mitigation may include monitoring conducted by ARRC at a frequency agreed to by ADF&G." The Piledriver Slough Mitigation Plan was created to assess impacts of the Northern Rail Extension Project-Phase 1.

### Need

A levee was put into place to alleviate blockage to spring flow flushing from the Tanana River into the Piledriver Slough due to construction of the new rail extension. With the construction of the levee, concerns were raised about the potential alteration in flow-rate because of the lacking ability of natural flushing of debris or ice build-up by spring flows. Resulting concerns include ice and log jams and beaver dams impeding fish passage. This study was created to assess the risk that such obstructions pose to fish passage.

### Objectives

The Alaska Department of Fish and Game (AKDFG) consults TVWA in action through a Memorandum of Agreement implementing fish monitoring within the Piledriver Slough located in the City of Salcha and the 23 Mile Slough. TVWA is charged to manage the Piledriver Slough Beaver Activity Survey program until to 2022, in which a final report will be submitted to AKDF&G and the Alaska Rail Road (AKRR). The report will compile results and conclusions drawn from outlined objectives and accomplishments achieved during the 10-year study.

### Methodology

The ten-mile section of the Piledriver Slough was divided into two sections for managing monitoring based upon distance from the levee site to the Bailey Bridge. These sections were the *Upper Piledriver* and *Lower Piledriver*. Upper Piledriver surveying began from the levee site and ended at the Old Valdez Trail road crossing. This section was surveyed by TVWA staff with the assistance of citizen scientists from the Salcha Elementary. Lower Piledriver surveying began from the Old Valdez Trail road crossing ended at the Bailey Bridge, adjacent to Eielson Airforce Base. This section was surveyed by TVWA staff. Undivided, the 23 Mile Slough site was located and surveyed in its entirety off of Old Eielson Farm Road. All surveys took place late spring, summer, and late fall, which exact dates dependent on staff and school availability.

For the study of Upper Piledriver, TVWA trained volunteers and students who acted as citizen scientist through a presentation and science curriculum on water safety, fish, plant and invertebrate identification, fish handling, water quality, invasive species, aquatic invertebrates and habitat assessment. Each child was equipped with a tool kit containing supplies and safety for the field surveying. Algae and aquatic plant identification education curriculum was added in 2014. Fish factsheets, tracks sheets, and more complex habitat assessments were added in 2015. Compasses and magnifying boxes were added to the curriculum in 2016. In 2017 we added curriculum on water velocity and flow.

*Equipment:* Equipment used in the study by TVWA staff were a Garmin GPS 62s, PentaxWGIII SR Adventure Proof GPS Camera, GoPro videocamera, Android telephone camera for capturing photos and videos to be used for analysis and reporting. GPS units were used for marking identified dams and lodges as well as geo-referencing photos.

*Water Quality Sampling:* Adopt-a-Stream water quality sampling protocol was used to record water quality at each Upper Piledriver Site. This protocol is detailed below:

Step 1: Perform a Hanna meter pre-sampling check with tap water. Using the pH 4 and 1413 conductivity standards provided, test your meter's accuracy. Turn on your meter. Place a small amount of the pH 4 standard into plastic cup marked "pH4 check" (just enough to cover the sensor). Take a pH reading and record the result. It should fall between 3.8 and 4.2. Rinse the meter in tap water and shake it gently to remove excess water. Then, place a small amount of the 1413 conductivity standard into the plastic cup marked "conductivity check" and take a reading. Note the conductivity level. It should fall between 1342 and 1484. Rinse the meter again in tap water and shake it gently to remove excess water before replacing the cap. The standards are safe to pour down the drain with a little tap water. DO NOT pour them into the stream.

<u>Step 2: Collect water sample:</u> A few yards away (preferable downstream or down current) from your exact sampling site, rinse the plastic bucket three times with stream water. Then go to your site and, facing upstream, lower the bucket gently into the water, and fill it to a level about 2 inches from the lip of the bucket. If you are working in very shallow water, do not disturb the bottom while collecting the sample.

Step 3: Measure pH and Conductivity with Hanna Meter: Turn on the meter. Hold it or clip it to the side of the bucket in the sample water for 5 minutes. Turn on the meter. Press SET/HOLD until it is in conductivity ( $\mu$ ) mode, wait 15 seconds, then record three (3) sequential readings for Conductivity at 15 second intervals. Press SET/HOLD until it is in pH mode and wait 15 seconds. Record three (3) pH readings at 15 second intervals. Finally, press SET/HOLD until it is in temperature mode and wait 15 seconds. Record three (3) water temperature readings at 15 second intervals. Turn the meter off. Put the cover back on the meter, making sure to moisten the pH sensor before doing so.

<u>Step 4: Record the air temperature:</u> Hang the air thermometer somewhere where it will not lean against any soled object and where it is protected as much as possible from direct wind and sunlight. The thermometer will take at least five minutes to equilibrate. It might take longer if it has to adjust for large changes in temperature. Recording the air temperature after you have completed the water quality sampling should ensure that the thermometer has had ample time to adjust.

<u>Step 5: Perform the meter post-sampling check in office with tap water:</u> Using the pH 10 and 1413 conductivity standards provided, test your meter's accuracy. Turn on your meter. Place a small amount of the pH 10 standard into plastic cup marked "pH10 check" (just enough to cover the sensor). Take a pH reading and record the result. It should fall between 9.8 and 10.2. Rinse the meter in tap water and shake it gently to remove excess water. Then, place a small amount of the 1413 conductivity standard into the plastic cup marked "conductivity check" and take a reading. Note the conductivity level. It should fall between 1342 and 1484. Rinse the meter again in tap water and shake it gently to remove excess water before replacing the cap. The standards are safe to pour down the drain with a little tap water. DO NOT pour them into the stream.

*Fish Sampling:* Chena Salmon sampling protocol was used for recording information on fish. Sampling procedures follow. Gee-type minnow traps (23 x 45 cm, 0.64 cm wire mesh, with 2.5 cm diameter openings) will be baited with salmon roe and set 5-10 mm apart for a 24-hour soak time (Swales, 1987). After the 24 hour soak, volunteers will identify and count all fish in the trap and, for each Chinook salmon and Arctic lamprey, will determine weight using water displacement and length using a Photarium viewing box (Duvall, WA, USA) to estimate the condition, or K factor (Weatherly and Rogers 1978). Fish will be released after identification and measurements are taken. Any incidental fish deaths will be labeled and brought to the USFWS laboratory in Fairbanks for further processing.

### 1. Set Traps:

- Place bait ball in the trap
- Put trap in suitable location length-wise to current. Slow moving water with in-stream cover is best but this may not be possible at all sites. Put traps in the slowest moving water available at your site because fish will get exhausted swimming against current
- Let your trap soak overnight and check on it 24 hours later
- Be as consistent as possible with length of soak -me!
- Get traps in deep enough water to cover the trap (deeper is better)

- Don't put traps in a high use area because they may get vandalized or stolen
- Make sure that traps are well-secured to something on the bank

## 2. Checking Traps

- Have all of your equipment ready before removing any traps from the water.
- Fill your counting and holding buckets half full of river water.
- Remove one of your traps from the water and gently pour fish into your counting buckets.
- Catch one fish at a time with the dip net and place it in the viewing box to identify it.
- Go to your guide. If the fish has an adipose fin, use the upper key. If it doesn't have an adipose fin,

use the bottom key. Pictures & descriptions for each species are in the guide (with TVWA).

- Record length of first 10 fish you identify for each habitat type using length markings on viewing box or measuring tube.
- After identification, put fish into the holding bucket.
- After you are finished counting and identifying all of the fish from one trap gently pour the holding bucket into the river and start counting your next trap
- Record total numbers for each species on the datasheet if no fish are caught record that
- Complete one data sheet for both habitat types, try to keep neat, organized notes

<u>3. Fish Handling Guidelines- Our goal is to minimize stress, limit handling, treat them with respect!</u>

- Keep your hands wet at all times.
- Use bare hands, gloves can damage scales.
- Handle fish as little as possible.
- Only empty one trap into the counting bucket at a -me (to maximize oxygen content).
- If counting is taking a long -me you can try carefully changing out some of the water to maintain oxygen content and water temperature.
- Release fish in the same place where you caught them.

*Beaver Survey:* Beaver dams and lodges were surveyed visually by foot on Upper Piledriver Slough and by canoe on Lower Piledriver. Beaver dams were defined as dams built by beavers to provide ponds as protection against predators such as coyotes, wolves, and bears, and to provide easy access to food during winter. Beaver lodges were defined as dwellings constructed on the side of the stream that do not impeded passage. All dams and lodges were photographed, GPS locations were recorded, sites were described. Dams were measured for height, diameter of logs and width of passage. Dams were categorized based on activity by beavers (active, inactive) and type of dwelling (primary dam, secondary dam, lodge). Active was defined as dams or lodges that exhibited signs of recent activity including fresh chews, moved materials, feed piles, tracks, beaver slides, or beaver presence ect. Inactive dams and lodges were defined as places which did not exhibit the signs of use identified in the "active" definition. Primary dam was considered the largest dam in a  $\frac{1}{2}$  mile area that displayed the most use. Secondary dam was determined as a smaller dam.

## Follow Up:

All equipment was inventoried, cleaned, and serviced before and after the surveying season. Fish data reports were sent to the Alaska Dept of Fish and Game, in compliance with our permit requirement.

## **Study Survey Results**

TVWA contractor Heather Mirczak met with Salcha staff to discuss curriculum development in the spring. She took feedback from the teachers and discussed future ideas for the partnership and curriculum. The Piledriver project maintained strong community involvement throughout the study duration: 12 members of the Salcha Elementary School staff, 13 community and parent volunteers, 74 children attending Salcha Elementary School, 1 TVWA staff, 3 contractors, 5 volunteers, the Department of Fish and Game, and the U.S. Department of Fish and Wildlife.

The study had a total of twenty-eight survey sites. Eight survey sites (with 2 traps each) on the Upper Piledriver were within the periods of May 10-11, June 21-22 and August 18-19. This Upper Piledriver was monitored with the assistance of the Salcha Elementary School through the citizen scientist collaboration. Sixteen sites (with one trap each) were surveyed on Lower Piledriver by TVWA field technicians and volunteers on May 10-11, June 21-22 and August 18-19. Undivided, 23 Mile Slough had four survey locations that took place on May 13-14 by TVWA field technicians and volunteers. TVWA staff included supervision by Bryn McEloroy. Contractors were Jenna Jonas, Heather Mirczak, and David Jonas. Volunteers included Ellen Martin, Natalie Ott Schudt, Charlie Schuldt, Ole Schudt, Garrett Schuldt, and Christy Everett.

In 2015, TVWA staff began recording qualitative data after each float and continued this practice in 2017.

*Fish:* AKF&G issued TVWA a Fish Resource Permit for the study (See Appendix A). Surveying took place post-permit issuance. Data collection recorded fish species identified, relative size, and location assisted by equipment (minnow traps, viewer, bucket, and identification book). The compilation of fish parameters was reported to AKRR as the *Fish Collection Report* (See Appendix B). Fish monitoring was conducted at 28 sites with a total of 40 caught-and-release fish recorded.

*Beaver:* Beaver dams were categorized based on whether or not it was actively used by beavers, which simply were active or inactive. Secondary categorization was based on dwelling type of dam, which consisted of primary dam, secondary dam and lodge. Dam activity and dwelling type was recorded as well as coordinates.

## **Discussion of Study Outcomes & Activities**

Successful implementation of the Piledriver Sough Project 2017 provided survey data recording and community buy-in through community-based learning as citizen-scientist volunteers ranging from youth or adults. TVWA featured the Piledriver project at several outreach events including the Salcha Open House/Personalized learning launch on September 19 2017. A display provided information to the public that outlined the full scope of the Piledriver Project and highlighted the Salcha Elementary School children's stewardship accomplishments. Piledriver curriculum was used at other TVWA water sampling events such as those used with the Adopt-A-Stream program.

## **Hydrology Monitor**

2017 was not a scheduled year for hydrology monitoring, thus there is no data recorded in 2017.

## **Appendix A: Fish Resource Permit: Fish Resource Permit**



STATE OF ALASKA DEPARTMENT OF FISH AND GAME 333 Raspberry Road ANCHORAGE, ALASKA 99518

Permit No. SF2017-100

Expires: 10/15/2017

FISH RESOURCE PERMIT (For Scientific/Collection Purposes)

This permit authorizes:

<u>Jenna Jonas</u> (whose signature is required on page 2 for permit validation)

 Tanana Valley Watershed Association

 516 Second Avenue, Suite 412, Fairbanks, AK 99701

 (248) 568-0345

 jenna.tvwa@gmail.com

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to conduct the following activities from May 1, 2017 to October 15, 2017 in accordance with AS 16.05.930 and AS 16.05.340(b).

Purpose: To examine fish presence and abundance in the target locations in fulfillment of Mitigation Measure 56 of the Service Transportation Board.

Location: Piledriver slough (334-40-11000-2490-3315), 23 mile slough (334-40-11000-2490-3315-4010)

Species: Local species

Method of Capture: Minnow trap (Stipulations #5-7)

Final Disposition: Any number of fish may be captured, identified, and quickly released alive during each sampling event.

≤50 individuals each of Alaskan brook lamprey, Arctic lamprey, and Chinook salmon at each sample location may also be measured before release.

≤2 individuals of each unknown species may be killed and saved for later identification.

All unintended mortalities must be recorded and may either be returned to capture site waters or provided to the U.S. Fish & Wildlife Service as vouchers.

COLLECTION REPORT DUE <u>November 15, 2017</u> and RESEARCH REPORT DUE <u>April 15, 2018</u>; see Stipulations #2 and #3 for more information. Data from such reports are considered public information. Reports must be submitted by email (<u>dfg.dsf.permitcoordinator@alaska.gov</u>) or by mail to: Alaska Department of Fish and Game, Division of Sport Fish-HQ, 333 Raspberry Rd, Anchorage, AK 99518. A report is required whether or not collecting activities were undertaken.

#### **GENERAL CONDITIONS, EXCEPTIONS, AND RESTRICTIONS**

- 1. This permit must be carried by person(s) specified during approved activities who shall show it on request to persons authorized to enforce Alaska's fish and game laws. This permit is nontransferable and will be revoked or renewal denied by the Commissioner of Fish and Game if the permittee violates any of its conditions, exceptions, or restrictions. No redelegation of authority may be allowed under this permit unless specifically noted.
- No specimens taken under authority hereof may be sold, bartered, or consumed. All specimens must be deposited in a public museum or a public scientific or educational institution unless otherwise stated herein. Subpermittees shall not retain possession of live animals or other specimens.
- The permittee shall keep records of all activities conducted under authority of this permit, available for inspection at all reasonable hours upon request of any authorized state enforcement officer.
- Permits will not be renewed until detailed reports, as specified in the Stipulations section, have been received by the department.
   UNLESS SPECIFICALLY STATED HEREIN, this permit does not authorize the exportation of specimens or the taking of specimens outside of existing regulations.

tamen ermit Coordinator

Permit Coordinator Division of Sport Fish

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Director Division of Sport Fish

3-10-17

Date

#### SF2017-100 continued (page 2 of 2)

Authorized Personnel: The following persons may perform collecting activities under terms of this permit:

#### Jenna Jonas, David Jonas, Bryn McElroy, Christie Everett, Heather Mirczak, Annie Keep-Barnes, Torri Brannan, Ben Kennedy

Employees and volunteers under the direct supervision of, and in the presence of, one of the authorized personnel listed above may participate in collecting activities under terms of this permit.

#### Permit Stipulations:

- Klaus Wuttig (459-7344; <u>klaus.wuttig@alaska.gov</u>), the Tanana River Area Management Biologist (AMB), must be contacted for final authorization **prior** to you engaging in any collecting activities. <u>The time/date of this contact must be included in your</u> <u>collections report (using the "data submission form" furnished by ADF&G</u>). AMBs have the right to specify methods for collecting, as well as limiting the collections of any species by number, time, and location.
- 2) A report of collecting activities, referencing this fish resource permit, must be submitted within 30 days after the expiration of this permit. The report (using a data submission form furnished by ADF&G), shall include all species, numbers, dates, locations of collection (datum/GPS coordinates in the decimal degrees format (dd.ddddd)), and disposition, and if applicable, sex, age, and breeding condition, and lengths and weights of fish handled. It must also include the date/time the local biologist was contacted for final authorization to carry out collecting activities.
- 3) A report of research activities, referencing this fish resource permit, must be submitted within 6 months after the expiration of this permit. This report should present the research conducted in a format similar to a scientific paper including the following: introduction (objective of the study plan and hypothesis), methods, and results. The report is intended to show that the specimens were used in a scientific method, and allows for the evaluation of potential cumulative effects from multiple projects in the same area. A report is required whether or not collecting activities were undertaken.
- 4) An instance of >10% unintended collecting mortality requires sampling at a site to cease and the AMB contacted.
- 5) Each piece of unattended sampling gear must be: 1) labeled with the permittee's name, telephone number, and permit number, 2) properly secured to ensure retrieval, 3) placed in a location where they will not be easily noticed (e.g. under cut banks, in pools away from roads or trails), 4) allowed to soak no more than twenty-four hours at a time, 5) located with GPS coordinates, and 6) accounted for and removed at the conclusion of sampling.
- 6) Salmon eggs used as bait in traps must either be sterilized commercial eggs or, if raw, disinfected prior to use. A 10-minute soak in 1/100 Betadyne solution or some other iodophor disinfectant is adequate. Commercial eggs must be placed into a container that does not allow the fish to consume them (e.g., film canister with holes punched in it or a perforated plastic bag).
- 7) Gloves, boots, and collecting gear should be cleaned and disinfected between streams to reduce the potential of pathogen transmission and the spread of invasive species. Clean all equipment free of sediment, vegetation, and seeds, and then wash/rinse in 1/100 Betadyne solution or soak in 10% bleach solution for a minimum of 10 minutes. Felt or absorbent soles on waders and wading boots are prohibited.
- 8) If new anadromous fish species <u>or</u> previously undocumented life stages of anadromous fish are found in permitted streams, rivers, and lakes, the permit holder will work closely with ADF&G to see that information is included in the database for the Catalog of Waters Important for Spawning, Rearing or Migration of Anadromous Fishes. Anadromous fish include Oncorhynchus spp., Arctic char, Dolly Varden, sheefish, smelts, lamprey, whitefish, and sturgeon. Please direct questions to J Johnson (907-267-2337; j.johnson@alaska.gov).
- 9) Contact Tammy Davis with the ADF&G Invasive Species Program (907-465-6183 or 1-877-INVASIV), and the nearest AMB (Stipulation #1) within 24 hours should you find any species suspected to be a non-native species during your sampling. If possible the organism should be killed, preserved by freezing or placing into 90% alcohol, and taken to the nearest ADF&G office. Please take a photo of the organism, as well as a photo of the organism in the environment in which it was observed, and note the location with a GPS or by describing it on a map with landmarks.
- 10) No fish may be possessed live or transported live without a valid Fish Transport Permit (FTP) obtained from the Alaska Department of Fish and Game.
- 11) A copy of this permit, including any amendments, must be made available at all field collection sites and project sites for inspection upon request by a representative of the department or a law enforcement officer.
- 12) Issuance of this permit does not absolve the permittee from securing any other required state, federal, or local permits, including securing permissions to trespass on controlled lands.
- 13) Failure to comply with the conditions of this permit will result in the loss of future permitting privileges.
- 14) PERMIT VALIDATION requires permittee's signature agreeing to abide by permit conditions before beginning collecting activities:

Jain Herto

Signature of Permittee

ecc: Klaus Wuttig, Division of Sport Fish, Fairbanks Brandy Baker, Division of Sport Fish, Delta Junction Bonnie Borba, Division of Commercial Fisheries, Fairbanks Audra Brase, Division of Habitat, Fairbanks Michelle Morris, Commercial Fisheries Permit Coordinator, Juneau Colonel Bear, Alaska Wildlife Troopers Captain Leath, Alaska Wildlife Troopers Northern Detachment

# Appendix B: Fish Collection Report: Fish Collection Report

## **Summary**

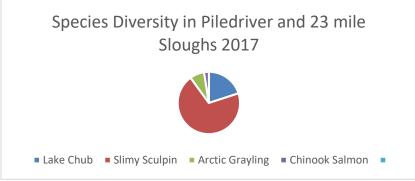
In 2017 40 fish were caught, identified and released in piledriver and 23 mile sloughs. Of these, 13 were caught on Upper Piledriver with the Salcha Elementary students and 23 on Lower Piledriver, and 4 on 23 mile slough.

Place	Total Fish Caught	Slimy Sculpin	Lake Chub	Burbot	Arctic Grayling	Chinook Salmon	Days Sampled	# traps set
Upper Piledriver	13	12	1	0	0	0	May 10- 11, June 21-22, August 18-19	16
Lower Piledriver	23	14	7	0	1	1	May 10- 11, June 21-22, August 18-19	16
23-Mile Slough	4	2	0	0	2	0	May 10- 11	4

## **Equipment Used**

Gee-type minnow traps (23 x 45 cm, 0.64 cm bar mesh, with 2.5 cm diameter opening) were baited with disinfected salmon roe and set for 24 hours for each sampling event. Traps were placed in a variety of habitat types including cut banks, slough mouths, in woody debris, and on either side of beaver dams. All captured fish were identified to species. The fork length of the fish identified at each site each week was measured using the ruler on a medium Photarium viewing box (Duvall, WA). Fish were released after identification and measurement.

## **Species Diversity**



The most commonly caught fish this year was the slimy sculpin (28 fish). We caught the majority of the slimy sculpin on upper piledriver in sites 1 and 2. The second most commonly caught fish was the lake chub (8). In past years, the Slimy Sculpin has been the most commonly caught fish in each year of sampling on Piledriver slough.

The slimy sculpin (Cottus cognatus), a bottom-dwelling fish, can be found throughout most of northern United States, Canada and Alaska. The slimy sculpin is an ambush predator. It feeds primarily on insects, but also eats crustaceans, fish eggs, and small fish. The slimy sculpins' size and poor swimming ability makes it a great prey item for larger fish. The slimy sculpin has been studied in waters where there is current acidification (water that is more acidic). The sculpin were found to be less active and have lower rates of reproduction when found in these waters. For these reasons, the slimy sculpin has been identified as a good indicator species (a species that indicates a change in environment by a difference in behavior or population size) for acidification in lakes and ponds and possibly for streams.<sup>1</sup> Our water quality data thus far does not show that acidification is occurring in Piledriver slough, the challenge that we have observed is an increased sedimentation. Interestingly, the Slimy Sculpin do fan their nests to remove silt, an adaptation that may allow them to thrive in the changing Piledriver Slough. The slimy sculpin moves to shallower waters during the spawning season, which is in the spring, usually after break-up. Males establish a nesting spot under a rock or log and groom the area by fanning fine sediment and moving small pebbles with its mouth out of the area. Males are territorial and can be aggressive towards other males. A male courts a female until she deposits her eggs, which are yellow to pink, on the underside of the rock or log. The female does this from the upside-down position while the male fertilizes the eggs with his milt. The female leaves after egg deposition. A single male may spawn with several females. Once the eggs are fertilized, the male guards his nest until the young fish are ready to leave. During this time the male fans the eggs to remove silt and provide oxygen and keeps the nest clean. The eggs hatch about 30 days after being fertilized. The sac-fry stay in the nest, usually resting on the bottom. They remain there for about a week while the yolk is being absorbed. Once the yolk-sac is gone, the sculpin leave the nest as fry.

The lake chub (Couesius plumbeus) belongs to the largest freshwater fish family, the minnows (Cyprinidae). They are a small fish, with adults averaging from 5-10 cm long. The lake chub is found in all types of freshwater bodies (lakes and streams), but in Alaska it has been found more often in silty waters. It tends to prefer shallow water, although it will move to deeper water during hot weather. The lake chub is usually abundant wherever it is found. Young lake chubs feed primarily on zooplankton. Older lake chubs feed on terrestrial and aquatic insects, but also feed on algae, occasionally small fishes, and have been known to scavenge on decaying fish.<sup>2</sup> This makes sense as our surveys of Piledriver's aquatic invertebrates have found the habitat to be host an extensive array of aquatic insects and we have observed increasing amounts of algae in the past years. We additionally believe that we encountered the Lake Chub during their spawning period, which is known to occur between spring and early summer. This would account, in part, for their abundance although lake chub prefer spawning areas with shallow water and rocky or gravelly bottoms.

TVWA field staff did not observed the large numbers of grayling, ranging in size from 2-16 inches traveling in schools, mostly heading upstream that we saw in 2015 and 2016. We observed some grayling, but much less than in years previous. In the August sampling, no grayling were observed

<sup>&</sup>lt;sup>1</sup> "Slimy Sculpin" Alaska Dept. of Fish and Game Wildlife Notebook Series, Kelly Mansfield, 2004

<sup>&</sup>lt;sup>2</sup> "Lake Chub" Alaska Dept. of Fish and Game Wildlife Notebook Series, Kelly Mansfield, 2004

upstream of the beaver dams, leading us to conclude that they dams were blocking passage. TVWA staff also noted large numbers of spawing chum salmon also downstream of the dams in the slough during the August 18-19 sampling on Lower Piledriver slough.

## **Number of Fish Caught**

In 2017 we caught 40 fish. In 2016 we caught 252 fish, in 2015 the total catch was 373 fish, in 2014 the total catch was 58, 2013 saw 24 fish and in 2012 101 fish were captured. The drop in fish numbers is due largely to the lack of lake chub. We believe this was because our dates were slightly different this year and we did not fish while the lake chub were spawning locally. There is not yet enough data to determine a significant trend in fish numbers.

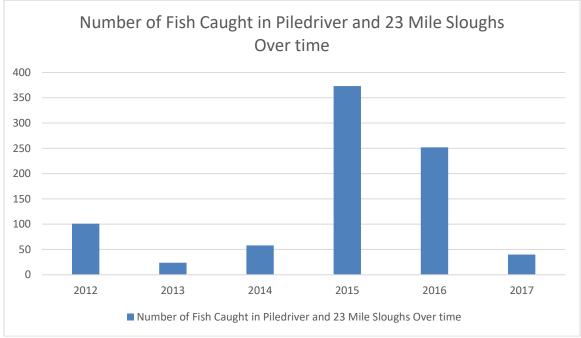


Figure 2: Number of Juvenile Fish Caught in Piledriver and 23 Mile Slough Annually

## **Sampling Sites**

The sites used in 2017 were, for the most part, consistent with those used in 2016.

## Table 2: Piledriver Slough site locations with Salcha School.

Upper Piledriver Sites 2017

			-
1	Culverts- Downstream	64.60180	147.09177
			-
2	Culverts- Upstream	64.60175	147.09187
3	Annie's Yard	64.60035	-147.0912
			-
4	Ingrid	64.59650	147.08459
			-
5	4-wheeler trail	64.59391	147.08321
			-
6	Xantheus Bridge	64.59293	147.07361
			-
7	Posted Braided	64.58728	147.06952
			-
8	Dam	64.58630	147.06807

## Table 3: Piledriver Slough site locations with TVWA staff and volunteers.

	Lower Piledriver Sites 2017		
	tied to willows just past culvert across from houses		
1	on River R	64.84387	147.71843
2	on island just past houses, river L	64.60297	147.08966
3	mealt frame river L	64.60259	147.0863
4	upstreamof cabin and camper, river L	64.60233	147.08534
5	donwstream of cabin/camper river R after old dam	64.60301	147.08369
6	downstream of giant old dam river R	64.60333	147.08543
7	upstream of second bridge dam River L	64.60429	147.08809
8	downstream in middle of bridge dam	64.60429	147.08809
9	upstream of 3rd dam bride river R by drift log	64.60547	147.08676
10	downstream of 3rd dam bridge by river R	64.60547	147.08676
11	On stick with pink flagging across from old lodge	64.60766	147.08507
12	Tied to dead snag in pool river L	64.60837	147.08905
10	set to stake out of beach, slough widens and path	(4 (1000	1 47 00007
13	flows right	64.61089	147.09007
14	on green twig upstream of gral island with cut spruce log, river L	64.61154	147.08916
15	in deep area between 2 shallow rocky ripples, river R	64.61420	147.08896
16	just past rocky ripplesbefore deep pool, river R at left V of river on left for before yellow tag on tree	64.61451	147.08911
17	river R	64.62028	147.0909
18	on right of Y after big log jam, river L	64.62366	147.08734

## **Appendix C: Beaver Report**

### Piledriver Slough Beaver Activity Survey Report 2017 Tanana Valley Watershed Association October 1, 2017

The Piledriver slough mitigation plan monitors changes to the Piledriver slough that may be caused by beaver activity. Due to construction of the new rail extension, a levee was put in place that blocks flushing flows into the Piledriver Slough from the Tanana River. The flow-rate changes may cause ice and log jams that would hinder fish passage. Beaver dams may no longer be knocked out by flushing spring flows and could cause further fish passage issues. Beavers are a natural part of the local environment and can help or hinder the other wildlife in the area. In the case of Piledriver Slough monitoring will be conducted to evaluate the beaver dams and determine if they need to be removed to aid fish passage through the slough.

The ten mile section of Piledriver from the levee site to the Bailey Bridge was monitored in two sections: "Upper Piledriver" from the levee site to the Old Valdez Trail road crossing and "Lower Piledriver" from the Old Valdez Trail road crossing to the Bailey Bridge adjacent to Eielson Airforce Base. Piledriver Slough was monitored on May 10-11, June 21-22 and August 18-19 2017. Identification of dam, and lodges were marked with GPS Locations. Pictures and videos were taken for further comparison and review. Beaver dam activity was classified as active or inactive and labeled as a dam, secondary dam and lodge.

Active beavers were present on Piledriver slough in 2017. We saw sign (chew sticks, vegetation removed, old dams re-constructed etc) during June sampling and during August sampling the two old dam sites (2 and 3) were fully constructed and blocking fish passage.

In response to the dams, TVWA contractors contacted James Durst of the Alaska Dept of Fish and Game was also contacted in June. He said "The NRE Piledriver Slough Beaver Dam Mitigation Plan project area is Piledriver Slough between the levee and the Bailey Bridge site across from EAFB. From what I can tell, the two dams in question are in the stream reach downstream of the Old Richardson Highway culverts and upstream of the Bailey Bridge crossing, so are in the lower half of the mitigation plan project area. This lower reach receives surface flows from an anadromous slough in addition to the groundwater-based flow from the upper portion of Piledriver Slough. Your upstream dam is in essentially the same location as Dam 1 in ADF&G Sport Fish's removal studies in the late 1990s and early 2000s, and your lower dam ("Dam 1.5") is at the upper end of the impoundment attributed to Sport Fish's Dam 2 and Dam 3. Dams 1, 2, and 3 were documented as blocking Arctic grayling, and their breaching resulted in grayling use of upstream areas. Klaus Wuttig worked on the Piledriver Slough system back then (report attached) and is now the Sport Fish Tanana River Area Manager. If Dam 1 and Dam 1.5 are blocking fish passage at this time (and it appears they likely are), the mitigation plan calls for consideration of breaching them at midsummer (about now). I won't be able to talk to Klaus until at least July 5 (he's in the field until Thu, and I'm out Wed-Tue) for his recommendation on not breaching, breaching now, or waiting until fall as part

of the mitigation project. Regarding actions that Alan Cady may take on his own, he knows that he needs to get a nuisance beaver permit from the Division of Wildlife Conservation if he wants to remove the beavers. We also told him that, except in special circumstances, using hand tools only to breach beaver dams does not require a fish habitat permit so he is free to do so if he chooses. If power equipment or machinery are used, a fish habitat permit is needed. In either case, removed material needs to be deposited in an upland location."

TVWA contractors also contacted Mark Peterburs of the Alaska Railroad in June, he believed that the dams were outside of the railroad's permit requirements and that the Railroad should not participate in removal. TVWA contractor corresponded with Alan Cady, a local land owner who had gone to ADFG to inquire about removing the dam downstream of his home. Mr. Cady said that he intended to remove the dam. He did not return a call in August.

Because the location of the dams was so close (1/2 mile approximately) from the uppermost limit of the observed spawning area and because without removing the beavers, there was not much point to removing the dam, TVWA contractors made a small breach on August 19 in the dams. We will continue to correspond with ADFG, the Railroad and Mr. Cady about this dam issue in the future.

### Dam Reference:

**Site 1: LB151-** Man-made bridge found in 2015 at this site was not found in 2017. 64.60296, 147.0882

**Site 2: LB152-** Old dam with woody debris buildup. This is an old dam that has naturally built up sediment and woody debris so that it ranges from 1-2 feet in height. 64.60339, 147.08543

Site 3: LB153- Large dam, Height ranged 1-2.5 feet with no passage. 64.60551, 147.08684



Dam 2 in August 2017



Dam 3 in May



TVWA volunteers Natalie, Ole and Charlie Schuldt navigate beaver dam 3 in June



Dam 3 in August 2017



TVWA volunteer Ellen Martin checks fish trap below Dam 3 in August sampling.

Composition of Dam 3

# **Appendix D: Photos**



Salcha  $5^{\ensuremath{\text{th}}}$  grade works on new stream velocity curriculum



Salcha students at Ingrid site



Salcha kindergarten students at Annie's site



Salcha students estimate fish length.



Salcha students examine slimy sculipin



Chinook salmon found in May sampling



Heather Mirczak canoes through aufeis in May